

Isolation and Characterization of Bacteria from Bekasam-Fermented Fish (*Rasbora sp*) and Their Antibacterial Potential

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ABSTRACT

Bekasam is a traditional fermented fish product. This study aimed to isolate and identify the bacteria involved in the fermentation of Bekasam from Riau and evaluate the antibacterial potential of their ethyl acetate extracts. Bacterial isolation was conducted using the pour plate method with selective media (MRS agar-CaCO₃ 0.5%). Antibacterial activity was tested against Gram-positive bacteria (*Staphylococcus aureus* and *Enterococcus faecalis*) and Gram-negative bacteria (*Escherichia coli* and *Pseudomonas aeruginosa*) using the agar diffusion method, with chloramphenicol as a positive control. Four bacterial isolates were identified: *Lactobacillus sp1* (BAC1), *Lactobacillus sp2* (BAC2), *Bacillus sp1* (BAC3), and *Bacillus sp2* (BAC4). All ethyl acetate extracts from these isolates demonstrated antibacterial activity. The highest activity was observed from the *Lactobacillus sp1* extract against *E. coli*, with an inhibition zone diameter of 16.1 mm.

Keywords: Bekasam, fermentation, *Lactobacillus sp.*, *Bacillus sp.*, antibacterial

Introduction

Fermented foods result from controlled microbial growth and the transformation of food components through enzymatic action. Traditionally, the purposes of fermentation include food preservation and enhancement of sensory qualities. This process occurs spontaneously in a conducive environment (Melini et al., 2019), allowing native lactic acid bacteria (LAB) to thrive optimally in the fermented food. Numerous studies have demonstrated that microorganisms present in fermented foods confer health benefits and can reduce disease risk, making them viable as probiotics. Microorganisms isolated from fermented foods can also reduce enzymes, antibiotics, vitamins, and bioactive metabolites (Qian et al., 2020).

Bekasam is a traditional fermented fish product that is widely popular in various regions of Indonesia. Each region has its distinctive type of bekasam, determined by the type of freshwater fish used. The production process involves using salt and roasted rice, which stimulate the spontaneous growth of LAB. LAB and substrates play crucial roles in the fermentation process, leading to changes in aroma and texture and contributing to preservation, thus extending the final product's shelf life (Haryo et al., 2024).

Bioactive compounds derived from bacteria in fermented foods exhibit diverse bioactivities, particularly those sourced from LAB. For instance, *Lactobacillus plantarum* isolated from spontaneously fermented goat milk has demonstrated probiotic properties, including antimicrobial effects against several pathogenic bacteria (Moreno-Montoro et al., 2018). Another study identified antimicrobial compounds such as bacteriocin from bacteria isolated from Kimchi (Yi and Kim, 2023). (Li et al., 2021) reported that *Lactobacillus* isolated from fermented soybean milk produced antioxidant compounds.

Isolation and bioactivity testing of metabolite compounds from bacteria isolated from bekasam have been performed. Wikandari et al., 2013 demonstrated that *L. plantarum* isolated from bekasam exhibited angiotensin-converting enzyme (ACE) inhibitory activity. Additionally, (Dewanti et al., 2017) reported that

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Lactobacillus acidophilus isolated from bekasam produced bioactive components such as lovastatin and bioactive peptides capable of inhibiting cholesterol synthesis. *Pediococcus acidilactici* isolated from bekasam of Sepat fish showed antibacterial activity against several pathogenic bacteria (Melia et al., 2019).

Based on the literature review, there is limited information regarding the bacteria involved in fermented products, particularly in Riau's bekasam. Therefore, further research on this product is necessary, especially on the acid-producing bacterial isolates from bekasam and their antibacterial potential

Materials and Methode

Materials

Bekasam purchased from Pasar Jumat, Muara Lembu village, Singingi sub-district, Kuantan Singingi district, Riau province; 0.9% NaCl, De Man–Rogosa–Sharpe (MRS) agar (Merck®), Mueller Hinton Agar (MHA) (Oxoid®), CaCO₃, sterile distilled water, ethanol 96% (Brathachem®), ethyl acetate (Brathachem®), disk paper.

Equipment

Analytical balance (KERN®), laminar airflow (INNOTECH-BIOBASE®), autoclave (HIRAYAMA®), Erlenmeyer flask (AGC Iwaki®), test tubes (Borosil®), Petri dishes (Normax®), inoculating loop, beaker glass (AGC Iwaki®), micropipette (Socorex Acura®), incubator (Mettler®), Microscope (Olympus®), Hotplate (IKA®) were used in this study.

Methods

1. Isolation of Bacteria from Traditional Fermented Food (Bekasam)

The bacteria were isolated from bekasam using the pour plate method. One gram of bekasam sample was ground in a sterile mortar to obtain a homogeneous sample. The homogenized sample was diluted with 0.9% NaCl to a total volume of 10 ml, resulting in a 10⁻¹ dilution. From the 10⁻¹ dilution, 1 ml was taken and added to a test tube containing 9 ml of NaCl, resulting in a 10⁻² dilution, and so on until a 10⁻⁵ dilution was obtained. From the 10⁻⁴ and 10⁻⁵ dilutions, 1 ml each was taken and inoculated onto Petri dishes containing MRS agar medium (supplemented with 0.5% CaCO₃). The Petri dishes were incubated at 37°C for 48 hours. Different bacterial colonies or those showing a clear zone were aseptically taken with an inoculating loop and streaked onto MRS agar medium. They were incubated in an incubator at 37°C for 48 hours. Purification was repeated until pure bacterial isolates were obtained (Zahra et al., 2018).

2. Identification and Characterization of Bacterial Isolates from Traditional Fermented Food (Bekasam)

Bacterial isolates were identified and characterized macroscopically and microscopically at the Bukittinggi Veterinary Center through organoleptic observation, Gram staining, catalase, motility, and endospore tests.

3. Fermentation and Extraction of Secondary Metabolites from Bacterial Isolates

Two inoculating loops of bacterial isolates were inoculated into 25 ml of sterile distilled water and homogenized. The prepared bacterial suspension was inoculated into 225 ml of previously sterilized NB medium and then placed on a rotary shaker at 120 rpm at room temperature for 48 hours. The bacterial fermentation product was mixed with ethyl acetate in a 1:1 ratio, stirred, and left for 24 hours. The ethyl acetate phase was separated using a separating funnel and evaporated using a rotary evaporator at 60°C. The extract was weighed and added with 0.5 ml of dimethyl sulfoxide (DMSO) (Handayani et al., 2015).

4. Antibacterial Activity Test of Ethyl Acetate Extract

The method used for testing the antibacterial activity of the ethyl acetate extract of bacterial isolates was the agar diffusion method using the MHA agar medium. The test samples (0.2 g/mL) were ethyl acetate extracts of bekasam bacterial isolates dissolved in DMSO. The test bacteria used were Gram-positive bacteria, *Staphylococcus aureus* and *Enterococcus faecalis*, and Gram-negative bacteria, *Escherichia coli* and *Pseudomonas aeruginosa*. The bacterial suspension was prepared in a sterile 0.9% NaCl solution with

turbidity corresponding to McFarland standard 0.5. Ten μl of the sample suspension (ethyl acetate extract of bekasam bacterial isolate, chloramphenicol (positive control), DMSO (negative control) was dropped onto sterile disk paper, and each disk was placed on the medium of each test bacterium. Each test was performed in triplicate. The Petri dishes were incubated at 37°C for 48 hours. The clear zone was measured using a caliper (Efendi et al., 2022).

Data Analysis




Antibacterial activity was measured by observing the diameter of the inhibition zones on the disk paper, and the data were analyzed descriptively.

Result and Discussion

The bekasam product used in this study was purchased from Pasar Jumat, Muara Lembu village, Riau province. The study began with bacterial isolation by cultivating bacteria on MRS agar, a selective medium for lactic acid bacteria (LAB) growth. Isolation was performed using the pour plate method, adding CaCO_3 to the medium to distinguish acid-producing bacteria (LAB) from non-acid-producing bacteria. This differentiation is indicated by clear zones around the bacterial growth. This occurs because CaCO_3 reacts with the acid produced by LAB, forming soluble calcium lactate, which results in clear zones around the growing colonies, as similarly observed by Olivia (Putri and Kusdiyantini, 2018). Generally, the isolated bacteria appeared round and cream-white, with spreading growth.

Based on macroscopic observation and the formation of clear zones in their growth media, visually distinct isolates were re-inoculated onto other media and repeatedly purified until pure isolates were obtained. Purity was confirmed by observing single colonies that did not overlap. Four pure isolates were obtained and coded as BAC1, BAC2, BAC3, and BAC4, which were then subcultured on slant MRS agar. Each bacterial isolate was subsequently characterized macroscopically and microscopically at the Bukittinggi Veterinary Laboratory (Reference No.: 29014/PK.301/F4B.1/12/2021). The identification results for each isolate are presented in Table 1.

Table 1. Identification results of bacterial isolates derived from bekasam

Code of Bacteria	Isolate of Bacteria	Bacteria
BAC1		<i>Lactobacillus sp1</i>
BAC2		<i>Lactobacillus sp2</i>
BAC3		<i>Bacillus sp2</i>

BAC4

*Bacillus sp1*

Several studies have linked microbes resulting from fish fermentation to various microorganisms, including lactic acid bacteria, *Staphylococcus spp.*, *Bacillus spp.*, and yeasts (Belleggia and Osimani, 2023). This supports previous findings that the four bacterial isolates identified belong to the genera *Lactobacillus sp.* and *Bacillus sp.* Research on these microorganisms is primarily centered on their fermentative capabilities and probiotic benefits. Additionally, they are known for producing valuable metabolites that are applied in nutraceuticals, pharmaceuticals, commodity chemicals, and flavor and aroma compounds. Furthermore, during fermentation, lactic acid bacteria can generate antimicrobial agents such as organic acids, hydrogen peroxide, and bacteriocins, which effectively inhibit the proliferation of spoilage and pathogenic bacteria (Sharma et al., 2019).

The bacterial cultures underwent extraction using ethyl acetate in a 1:1 ratio. Ethyl acetate, a semi-polar solvent with no antimicrobial properties, effectively dissolves both polar and non-polar compounds. Subsequently, the ethyl acetate filtrate was evaporated using a rotary evaporator at 60°C to obtain the extracts. The concentrations of these extracts were ranked in descending order as BAC2, BAC3, BAC4, and BAC1, depicted in Figure 1.

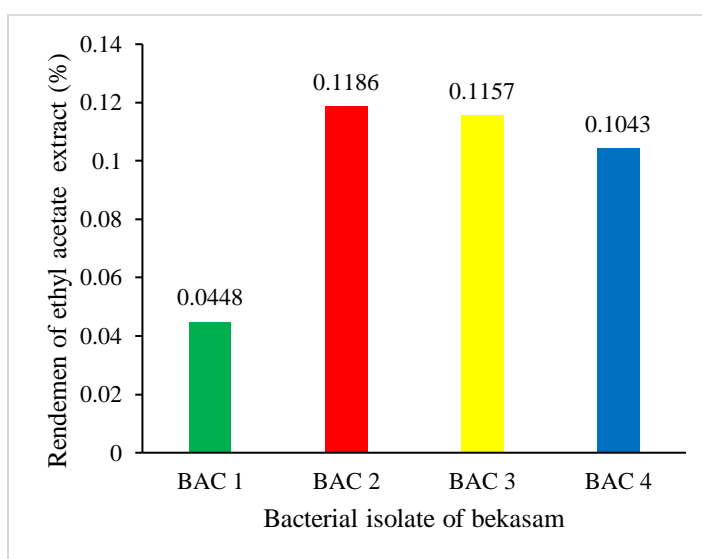


Figure 1. Yield of ethyl acetate extracts from bacterial isolates of Bekasam

Note: BAC1: *Lactobacillus sp1*; BAC2: *Lactobacillus sp2*; BAC3: *Bacillus sp2*; BAC4: *Bacillus sp1*

The antibacterial activity of the bacterial extracts was tested using the agar diffusion method with disk paper (Kirby & Bauer method). The positive control was 3 mg/ml chloramphenicol, a broad-spectrum antibiotic effective against Gram-positive and Gram-negative bacteria. Antibacterial activity was categorized based on the diameter of the inhibition zones: weak activity for diameters of 7-9 mm, moderate activity for 9-12 mm, and strong activity for ≥ 12 mm (Sharma et al., 2009). Four bacterial extracts exhibited antibacterial activity to *Staphylococcus aureus*. One isolate showed weak activity with an 8.1 mm diameter (BAC4), and Three isolates showed strong activity with diameters of 15.75 mm (BAC1), 13.45 mm (BAC2), and 12.2 mm (BAC3). Four extracts showed antibacterial activity to *E. coli*. Two isolates had strong activity with diameters of 16.1 mm (BAC1) and 15.7 mm (BAC2). BAC3 exhibited moderate activity with a 12.05 mm diameter, while BAC4 showed no antibacterial activity. Three extracts exhibited antibacterial activity to *Pseudomonas aeruginosa*. One isolate had moderate activity with a 9.65 mm diameter (BAC3), and two

isolates had strong activity with diameters of 13.1 mm (BAC1) and 13.45 mm (BAC2). BAC4 showed no antibacterial activity. Two extracts showed antibacterial activity against *Enterococcus faecalis*. Two isolates had moderate activity with diameters of 13.15 mm (BAC1) and 11.5 mm (BAC2). BAC3 and BAC4 showed no antibacterial activity.

Several studies have underscored the capability of *Lactobacillus sp.* to mitigate bacterial infections following oral administration in mice. Bacteriocins produced by probiotics are characterized as small, cationic, hydrophobic, and heat-stable peptides. Moreover, bacteriocins are primarily recognized for their potent antimicrobial activity against pathogens (Qian et al., 2020). These peptides operate through two antibacterial mechanisms: either by binding to lipid II to inhibit cell wall synthesis, thereby inducing cell death or by utilizing lipid II as a docking molecule to initiate membrane insertion and pore formation, resulting in swift cell death (Cotter et al., 2005).

Table 2. Antibacterial activity of ethyl acetate extracts from bacterial isolates

Ethyl Acetate Extract	Average Inhibition Zone Diameter (mm)			
	<i>S. aureus</i>	<i>E. coli</i>	<i>P.aeruginosa</i>	<i>E.faecalis</i>
BAC1	15.75	16.1	13.1	13.15
BAC2	13.45	15.7	13.45	11.5
BAC3	12.2	12.05	9.65	-
BAC4	8.1	-	-	-
K +	16.65	16.4	13.6	16.85
K -	-	-	-	-

Note: BAC1: *Lactobacillus sp1*; BAC2: *Lactobacillus sp2*; BAC3: *Bacillus sp2*; BAC4: *Bacillus sp1*

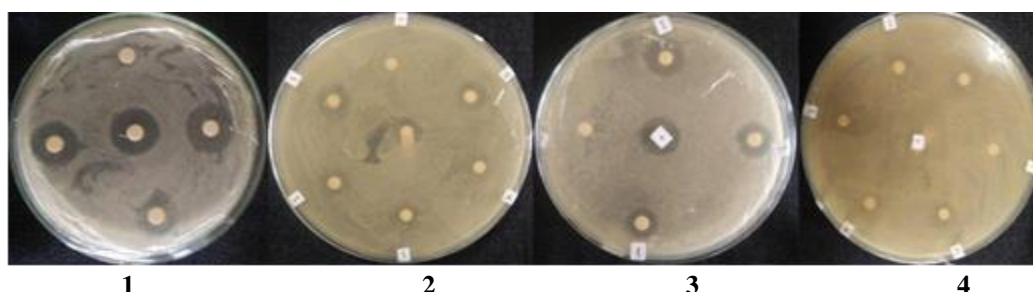


Figure 2. Antibacterial activity testing of ethyl acetate extracts from bacterial isolates of Bekasam

Note: 1: *Staphylococcus aureus*, 2: *Escherichia coli*, 3: *Pseudomonas aeruginosa*, and 4: *Enterococcus faecalis*

Conclusion

This research derived four bacterial isolates from fermented fish products (Bekasam). Two isolates were identified as members of the *Lactobacillus sp.* group (BAC1 and BAC2), while the other two were identified as belonging to the *Bacillus sp.* group (BAC3 and BAC4). All ethyl acetate extracts from these isolates demonstrated antibacterial activity, with the most potent antibacterial effect observed in the ethyl acetate extract from *Lactobacillus sp1*.

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